

REVIEW

Open Access

Isolated biomolecules of pharmacological interest in hemostasis from *Cerastes cerastes* venom

Fatah Chérifi¹ and Fatima Laraba-Djebari^{2*}

Abstract

Biomolecules from *Cerastes cerastes* venom have been purified and characterized. Two phospholipases isolated from *Cerastes cerastes* venom share 51% of homology. CC2-PLA2 exhibits antiplatelet activity that blocks coagulation. CCSV-MPase, a non-hemorrhagic Zn²⁺-metalloproteinase, significantly reduced the plasmatic fibrinogen level and hydrolyzes only its B β chain. Serine proteinases such as RP34, afa α cytin and CC3-SPase hydrolyze the fibrinogen and are respectively α , $\alpha\beta$ and $\alpha\beta$ fibrinogenases. In deficient human plasma, afa α cytin replaces the missing factors VIII and IX, and activates purified human factor X into factor Xa. It releases serotonin from platelets and directly aggregates human (but not rabbit) blood platelets. RP34 proteinase also had no effect on both human and rabbit blood platelet aggregation. CC3-SPase revealed a pro-coagulant activity. However, the insolubility of the obtained clot indicates that CC3-SPase does not activate factor XIII. In addition, CC3-SPase clotting activity was carried out with human plasmas from volunteer patients deficient in clotting factors. Results showed that CC3-SPase shortens clotting time of plasma deficient in factors II and VII but with weaker clotting than normal plasma. The clotting time of plasma deficient in factor II is similar to that obtained with normal plasma; suggesting that CC3-SPase is able to replace both factors IIa and VII in the coagulation cascade and thus could be involved in the blood clotting process via an extrinsic pathway. These results imply that CC3-SPase and afa α cytin could repair hemostatic abnormalities and may replace some factors missing in pathological deficiency. Afa α cytin also exhibits a fibrinase property similar to a plasmin-like proteinase. Despite its thrombin-like characteristics, afa α cytin is not inhibited by plasmatic thrombin inhibitors. The procoagulant properties of afa α cytin might have potential clinical applications.

Keywords: *Cerastes cerastes* venom, Proteinases, Phospholipases A2, Platelets, Blood-clotting, Hemostasis

Introduction

Serine proteases and phospholipases A2 isolated from snake venoms act on the hemostatic system as procoagulants, anticoagulants, pro- or anti-platelet aggregants. Some of these isolated molecules, mainly from *Viperidae* venoms, are used in diagnosis or treatment of thrombotic diseases and ischemic heart disease. Metalloproteinases can cause hemorrhage after accidental or experimental envenomation. However, some of these metalloproteinases are directly involved in the clotting of blood as they can act on fibrinogen and/or fibrin; they are called in this case fibrinolytic metalloproteinases. Their fibrinolytic activity makes them potent inhibitors of blood coagulation. Phospholipase A2 exert their anticoagulant effect by their

ability to inhibit platelet aggregation due to their high affinity to bind to activated factor Stewart (FXa). All these biological effects based on their direct involvement in hemostasis, let consider these molecules as potential tools or biomarkers in blood diseases.

Review

Viperidae and Crotalidae venoms are rich sources of hydrolytic enzymes and produce a complex pattern of clinical and toxic effects such as coagulation disorders, hemorrhage and necrosis [1-10]. Some of venom components act at various stages of the coagulation cascade. These components perform antagonistic functions, whilst some of them act synergistically. Therefore, the venom toxicity cannot be attributed to only one component [11]. However, most venom components produce beneficial effects when they act alone [12]. Snake venom also contains non-protein components including citrate,

* Correspondence: flaraba@hotmail.com

²USTHB, Faculty of Biological Sciences, University of Sciences and Technology Houari Boumedienne, BP 32 El Alia, Bab Ezzouar, Algiers, Algeria
Full list of author information is available at the end of the article

metal ions, carbohydrates, nucleotides as well as low concentrations of free amino acids and lipids [13-15].

Phospholipases A2 (PLA2s) represent more than 10% of the dry weight of the snake venoms from which they are isolated. PLA2s isolated from Viperidae venoms consist of 125–130 amino acid residues cross-linked by seven disulfide bonds which confer stability on the molecule and are calcium-dependant [16]. In addition to their hydrolytic activity, PLA2s may display many other activities, such as edematous, neurotoxic, cardiotoxic, hemolytic, convulsive, antiplatelet, antitumoral and anticoagulant properties. Based on their anticoagulant activity, PLA2s can be clinically useful against thrombotic diseases and for the diagnosis and treatment of hemostatic disorders [17]. The anticoagulant activity is due to their ability to inhibit platelet aggregation through factor Xa (FXa) blockade. Based on their direct involvement in the hemostatic cascade, PLA2 could also be used as tools or biomarkers in blood diseases.

Metalloproteinases found in Viperidae venoms may cause local hemorrhaging following accidental or experimental intradermal or subcutaneous injection of venom [18]. Some of them are known to display fibrinolytic activity. Fibrinolytic metalloproteinases dissolve fibrin clots and prevent clot formation by hydrolyzing fibrinogen, thus enhancing the toxic effect of hemorrhagic metalloproteinases, giving rise to pathological bleeding [19]. However, similar to PLA2s, metalloproteinases could be clinically useful against thrombotic diseases thanks to their potential use in laboratory tests or as therapeutic agents [20,21]. These proteinases may be useful for investigating the mechanisms of blood coagulation and platelet aggregation [11,12].

In addition to their beneficial effects, venom molecules are the cause of health problems after snake envenomation. Annually, more than 100,000 deaths are recorded worldwide, including 20,000 on African continent, while 400,000 victims retain severe and permanent functional sequelae [3]. Epidemiological data estimate envenomation cases at more than 5 million per year, with a mortality rate of 2.5%. In tropical Africa, Viperidae bites are responsible for 90% of envenomations [22]. Currently, the only available specific treatment against viper envenomation is immunotherapy [23-26]. Indications for this therapy, its mode of administration and type of antibody preparation in the form of F(ab')₂ or Fab still need to be elucidated [27-30]. Improvement of this treatment requires knowledge of the kinetic distribution of venom in vascular and tissue compartments.

Cerastes cerastes venom is a mixture of various proteins with broad biological and physiological activities; most of them are proteinases while some have been well characterized. Most of these proteins act on blood coagulation, including PLA2, the thrombin-like enzymes

RP34 and afaâcytin, anticoagulant protease fraction, aggregant serine protease, hemorrhagic metalloproteinase CcH1, and the non-hemorrhagic metalloproteinase CCSV-MPase [11,12,31-35]. In this paper, we report pharmacological activity of biomolecules isolated from *Cerastes cerastes* on hemostasis process.

Biochemical properties of biomolecules and their proteomic identification

Several molecules from *Cerastes cerastes* venom act on hemostasis, such as RP34, a serine proteinase which consists of two subunits of 48.5 kDa [32]. Another serine proteinase, a thrombin-like molecule denominated afaâcytin, was purified and characterized [11]. Afaâcytin presents caseinolytic, arginine-esterase and amidase activities. It is a homodimeric proteinase with two subunits, alpha and beta, which have the same apparent molecular mass (40.0 kDa for each unit) and are indistinguishable in the absence of reduction or/and deglycosylation [11]. Both α and β chains are N-glycosylated. The two chains present the same N-terminal sequence (20 residues) which is similar to the sequence of other proteinases isolated from snake venom.

Three molecules – CC2-PLA2, CCSV-MPase and CC3-SPase – were characterized by proteomic analysis. Results showed some sequence similarity with other homologous enzymes isolated from several venoms [11,12,17,31-34]. CC2-PLA2, another PLA2 found in the same venom, presents 51% sequence homology with a previously purified molecule from the same venom by Laraba-Djebari *et al.* [32] (accession number in NCBI is gi |129506|), *i.e.* 61 out of 120 amino acid residues are common to the two PLA2s [31]. The peptide sequence of the new PLA2 was obtained by alignment with sequences of other venom PLA2s. Some snake venom proteinases were identified sharing sequence homology with CCSV MPase, four of which belonged to the metalloproteinase family (Group III snake venom metalloproteinase and Zn⁺²-metalloproteinase disintegrin) [17]. Three of these proteins corresponded to *Cerastes vipera* venom and the others to venom proteins of different snake species [17].

All of these proteins were identified with at least two unique specific peptides and presented similarities with the purified CCSV-MPase. Previous studies showed that CCSV-MPase, characterized by SDS-PAGE analysis, could be classified as a member of the high-molecular-mass metalloproteinase family, due to its molecular mass estimated at 70 kDa in both reducing and non-reducing electrophoresis conditions [12]. Furthermore, the partial amino acid sequence of CCSV-MPase was identified by MALDI-TOF MS/MS analysis. Based on its molecular mass and partial amino acid sequence, CCSV-MPase may be classified in the P-III class of

SVMPs containing a disintegrin-like metalloproteinase, with cystein-rich domains.

It is well established that some venom components have beneficial effects when acting in isolation. *Cerastes cerastes* venom is a mixture of protein components with multiple actions including coagulation [11,32,36,37]. These proteins may induce hemorrhage and capillary permeability disorders, through their disintegrin domain or related proteins that disrupt primary hemostasis by acting on platelet adhesion. Thus, a single molecule can be endowed with several activities. The structural differences between proteins, natural factors of hemostasis, as well as the multiplicity of target components of the same venom, are elements that could explain the efficiency of partial immunotherapy [15]. Fibrinogenases (serine proteinases or metalloproteinases) are widespread in Viperidae venoms. They hydrolyze fibrinogen and/or degrade the fibrin clot, enhancing the effect of hemorrhagic metalloproteinases that give rise to pathological bleeding.

Coagulant and fibrinolytic activities of isolated molecules

Fibrinogen is a glycoprotein of 340 kDa with three polypeptide chains; A α (67 kDa), B β (50 kDa) and γ (43 kDa) linked by disulfide bonds. It can be hydrolyzed by thrombin, thus producing fibrin components and fibrinopeptides. Thrombin activity (control) on fibrinogen demonstrated the release of fibrinopeptide A (FpA) followed by fibrinopeptide B (FpB).

Proteolytic enzymes of *Cerastes cerastes* venom were identified as α , β or γ fibrinogenases depending on their ability to hydrolyze the fibrinogen *in vitro*. SDS-PAGE analysis of fibrinogen in the presence of venom revealed two entities (55 kDa and 50 kDa) indicating activities of α - and β -fibrinogenase. Purification and characterization of three procoagulant proteinases (RP34, afa α cytin and CC3-SPase proteinase) showed fibrinolytic activities when analyzed by SDS-PAGE, afa α cytin and RP34 displayed, respectively, α , β -fibrinogenase and α -fibrinogenase activity [11,32,34]. Like afa α cytin, CC3-SPase is also characterized as an α , β -fibrinogenase due to the release of both A and B fibrinopeptides.

Susceptibility of afa α cytin to diisopropyl fluorophosphate and benzamidine indicates the presence of a serine and an aspartic (or glutamic) acid residues in the catalytic site. Calcium is required for structural cohesion of the afa α cytin molecule [11]. CCSV-MPase cleaves only the B β chain of fibrinogen and exerted no action on A α or γ chains. This property contrasts with those of other SVMPs which preferentially cleave only the A α -fibrinogen chain. However, these metalloproteinases, belonging to the PI class of SVMPs, present low molecular mass, with only the metalloproteinase domain, as in

the case of fibrolase purified from *Akgistrodon contortrix contortrix*, piscivorase II of *Akgistrodon piscivorus piscivorus*, lebetase purified from the venom of *Vipera lebetina*, neuwiedase from *Bothrops neuwiedi* venom, the atroxase of *Crotalus atrox* and leucurolysins from venom of *Bothrops leucurus* [5,38-42].

Proteinases (afa α cytin, RP34, CC3-SPase and CCSV-MPase) showed caseinolytic activity as crude venom. CC3-SPase displayed arginine ester hydrolase activity while the CCSV-MPase does not. Both molecules presented a high amidolytic activity similar to that of crude venom. Previous results revealed that the use of specific inhibitors for serine proteinases and metalloproteinases showed that CC3-SPase is a thrombin-like Ca²⁺-dependent serine proteinase. Afa α cytin isolated from the venom of *Cerastes cerastes* showed that Ca²⁺ is essential for its activity not only as a cofactor but can contribute to the stability or structural cohesion of the enzyme [11]. CCSV-MPase appears to be a zinc-dependent metalloproteinase given that metal chelators, EDTA and 1,10-phenanthroline completely inhibited its proteolytic activity, which also suggested that unlike CC3-SPase, Ca²⁺ is not required for its catalytic activity. The sensitivity of the serine proteinase CC3-SPase, a specific inhibitor of thrombin (heparin and antithrombin III) may indicate that the receptor of CC3-SPase is identical to that of thrombin. Given its procoagulant properties and insensitivity to thrombin-specific plasma inhibitors, afa α cytin might be interesting to employ as a hemostatic agent in some types of hemorrhage, such as post-operative thrombocytopenia [11].

Serine-proteinases hydrolyze fibrinogen by acting on the two chains, α and β , of this substrate thereby causing the formation of a fragile fibrin clot. CCSV-MPase acts on the β chain of fibrinogen resulting in the release of only fibrinopeptide B. Afa α cytin, as a component of the venom (2% w/w), hydrolyzes fibrinogen in the same manner that CC3-SPase degrades firstly the A α chain and then, 24 hours later, the B β chain, leading to a fragile clot, which suggests that CC3-SPase, similarly to afa α cytin, is unable to activate the factor XIII responsible for the resistance of the fibrin clot [11,34]. CCSV-MPase cleaves only the B β chain of fibrinogen and exerts no activity on A α and γ chains. CCSV-MPase properties may allow its use as a therapeutic agent in some pathologies that require anticoagulant administration.

Most thrombin-like enzymes (TLE) isolated from snake venoms act on fibrinogen by hydrolyzing one chain rather than two, although the cleavage site is the same (Arg16-Gly17) as the α chain (Arg15-Gly16) on β chain, by releasing fibrinopeptides A or B as CCSV-MPase which degrades only the β chain of fibrinogen. CC3-SPase shortened the clotting time of plasma deficient in factor VII and II with a weaker clot than that formed with normal plasma. The clotting time of plasma

deficient in factor II is similar to that obtained with normal plasma after the action of serine protease CC3-SPase, which suggests that this molecule is able to replace factors IIa and VII.

Procoagulant and anticoagulant snake venom components often act at later stages of the coagulation cascade. The main targets of these components are fibrinogen, prothrombin, factor X and platelets [3,5,11,12,17].

Several molecules have been purified from Viperidae and Crotalidae venoms and characterized as FX activator factors that are used as biomarkers in many hemostatic disorders. Indeed, thrombin-like components serve as structural models to extend our understanding of the structure-function relationships of blood coagulation factors, some of which have been clinically used for the treatment of thrombotic diseases, and are employed as tools in clinical assays [43].

Combination of gel filtration and ion-exchange chromatography proved to be successful in obtaining milligram quantities of new pure TLEs from the venoms of *Crotalus durissus terrificus* and *Crotalus durissus collilineatus* [43]. Functional characterization indicates that both enzymes preferentially degrade the B β chain of bovine fibrinogen and present edema-inducing and coagulant activities. However, the TLE from *Crotalus durissus collilineatus* venom showed twofold higher coagulant activity with a minimum coagulant dose (MCD) of 0.6 $\mu\text{g}/\mu\text{L}$, whereas the enzyme isolated from *Crotalus durissus terrificus* indicated an MCD of 1.5 $\mu\text{g}/\mu\text{L}$ [43].

Recently, a TLE denominated BpSP-I was isolated from *Bothrops pauloensis* snake venom; its biochemical, enzymatic and pharmacological characteristics were determined. BpSP-I showed high clotting activity upon bovine and human plasma and was inhibited by PMSF, benzamidine and leupeptin. Moreover, this enzyme showed stability when examined at different temperatures (-70 to 37°C), pH values [3-9] or in the presence of divalent metal ions (Ca²⁺, Mg²⁺, Zn²⁺ and Mn²⁺). BpSP-I showed high catalytic activity upon substrates, such as fibrinogen, TAME, S-2238 and S-2288. It also showed kallikrein-like activity, but was unable to act upon factor Xa or plasmin substrates [44].

Reducing blood viscosity is often required in the treatment of thrombotic and ischemic heart diseases. Defibrinogenation of the plasma by some enzymes from snake venoms is of interest. Indeed, all of these defibrinogenating biomolecules sharing these properties could be used as tools in clinical applications and in basic research. Further studies, in pharmacology and toxicology should be undertaken to determine their mode of action *in vivo*.

Effect of molecules on platelet function

Biological characterization of CCSV-MPase and CC2-PLA2 has been shown to be highly anti-aggregative in

relation to human platelets. The antagonistic effect of CC3-SPase is of interest in the context of the antiplatelet action of the hemostatic system, and may be an effective tool for reducing blood viscosity, a property that is often necessary in the treatment of thrombotic diseases and ischemic heart syndrome due to platelet aggregation. Previous studies have already demonstrated that afaâcytin may replace the missing factors VIII and IX in deficient plasmas, and activate purified human factor X into factor Xa [11]. It releases serotonin from platelets and directly aggregates human (but not rabbit) blood platelets. On the other hand, RP 34 has no effect on platelet aggregation [32]. Several anticoagulant PLA2s from snake venoms have been isolated and well characterized. Recently, two phospholipases, known as CC-PLA2-1 and CC-PLA2-2 with antiplatelet aggregation activity, were isolated from *Cerastes cerastes* venom [45]. An anticoagulant PLA2 was isolated and characterized as an inhibitor of the prothrombinase complex through its specific binding to FX [46]. Ammodytoxin A (Atxa) and its natural ammodytoxin isoform C were isolated from *Vipera ammodytes ammodytes* venom and belong to group IIA secreted phospholipases. These two isoforms differ only by two amino acid residues (Phe 124 > Ile and Lys128 > Glu), but there are significant differences in toxicity. The mechanism by which they block coagulation has been elucidated. Complementary experiments using surface plasmon resonance showed complete inhibition of binding to FXa through calmodulin (CaM). The crystal structure showed that the C-terminal region required for binding to FXa and CaM is highly exposed and accessible for interaction with receptor proteins in the monomeric and dimeric forms of ammodytoxin [45].

Conclusion

Viperidae venoms, considered to be one of the most important bioresources, include pharmacologically active molecules such as proteinases (metalloproteinases and serine proteinases) and phospholipase A2 [46,47]. All of these molecules are of interest in biotherapy as biomedicines or may be used as diagnostic tools. Proteases and PLA2 act on the hemostatic system as procoagulants, anticoagulants, and as agents of pro- or anti-platelet aggregation. Some of these molecules, especially those isolated from Viperidae venoms, are used in the diagnosis and treatment of thrombotic and heart diseases. Some components act synergistically at different stages of the coagulation cascade [48].

Constituents of Viperidae venoms contain two categories of components that act antagonistically through activation or inhibition of coagulation factors and platelet aggregation. These compounds, able to hydrolyze the coagulation factors with high specificity, are divided into serine proteinases and metalloproteinases. Phospholipases

also display potent inhibition of platelet aggregation. Biomolecules of snake venoms are of great fundamental diagnostic and therapeutic interest. Therapeutically, proteinases from Viperidae venoms are widely used as anticoagulants. Furthermore, they are valuable tools for understanding the different mechanisms of hemostasis and are also used in the diagnosis of dysfunctions related to coagulation factors such as enzyme activity in thrombin-like venoms that are used for the fibrinogenopathy screening. Venoms are also used for diagnostic analysis of various coagulation factors (factors V, VII, X, platelet factor III, protein C and factor of Willbrand). Snake venom proteases are useful tools for studying coagulation reactions.

Ethics committee approval

The present study was approved by the Ethics Committee.

Competing interests

The authors declare no conflicts of interest.

Authors' contributions

Both authors collaborated in this work; they read and approved the final manuscript. FLD carried out the purification and characterization of Serine proteinases; RP34 and Afaâcytin and drafted the manuscript. FC with contribution of FLD, purified and characterized the molecules (CC2-PLA2, CC3-SPase and CCSV-MPase).

Acknowledgments

This work was partially supported by PNR and ANDRS (National Agency for the Development of Health Research, Oran, Algeria) projects. We are grateful to Benayad Tahar for technical assistance (Laboratory of Food Security and the Environment, Scientific Police, Algiers, Algeria), Abdelkader Namane and Jean-Claude Rousselle (Plateforme of Proteomics, CNRS URA 2185, Pasteur Institute of Paris) for proteomic analyses.

Author details

¹USTHB, Faculty of Biological Sciences, Laboratory of Cellular and Molecular Biology, BP 32 El-Alia, Bab Ezzouar, Algiers, Algeria. ²USTHB, Faculty of Biological Sciences, University of Sciences and Technology Houari Boumedienne, BP 32 El Alia, Bab Ezzouar, Algiers, Algeria.

Received: 18 September 2012 Accepted: 19 November 2012

Published: 1 May 2013

Reference

- Rivière G, Bon C: Immunothérapie antivenimeuse des envenimations ophidiennes: vers une approche rationnelle d'un traitement empirique. *Ann Inst Pasteur* 1999, **10**(2):173–182.
- Chippaux JP, Goyffon M: Venoms, antivenoms and immunotherapy. *Toxicon* 1997, **36**(6):823–846.
- Braud S, Bon C, Wisner N: Snake venom proteins acting on haemostasis. *Biochimie* 2000, **82**(9–10):851–859.
- Abib H, Laraba-Djebari F: Effect of 60Co gamma radiation on toxicity and haemorrhagic, myonecrotic and edema-forming activities of *Cerastes cerastes* venom. *Can J Physiol Pharmacol* 2003, **28**(12):1141–1147.
- Castro HC, Zingali RB, Albuquerque MG, Pujol-Luz M, Rodrigues CR: Snake venom thrombin-like enzymes: from reptilase to now. *Cell Mol Life Sci* 2004, **61**(7–8):843–856.
- Castro HC, Rodrigues CR: Current status of snake venom thrombin-like enzymes. *Toxin Rev* 2006, **25**(3):291–381.
- Kornalik F: Toxins affecting blood coagulation and fibrinolysis. In *Handbook of toxicology*. Edited by Shier WT, Mebs D, Dekker M. New York; 1990:697–709.
- Liu S, Sun MZ, Greenway FT: A novel plasminogen activator from *Agkistrodon blomhoffi ussurensis* venom (Abusv-Pa): purification and characterization. *Biochem Biophys Res Commun* 2006, **384**(4):1279–1287.
- Hamza L, Girardi T, Castelli S, Gargioli C, Cannata S, Patamia M, et al: Isolation and characterization of a myotoxin from the venom of *Macrovipera lebetina transmediterranea*. *Toxicon* 2010, **56**(3):381–390.
- Hamza L, Gargioli C, Castelli S, Rufini S, Laraba-Djebari F: Purification and characterization of a fibrinolytic and hemorrhagic metalloproteinase isolated from *Vipera lebetina* venom. *Biochimie* 2010, **92**(7):797–805.
- Laraba-Djebari F, Martin-Eauclaire MF, Mauco G, Matchot P: Afaâcytin and α , β -fibrinogenase from *Cerastes cerastes* (horned viper) venom, activates purified Factor X and induces serotonin release from human blood platelet. *Eur J Biochem* 1995, **233**(3):756–765.
- Chérifi F, Rousselle JC, Namane A, Laraba-Djebari F: Zn²⁺: a required ion for procoagulant metalloproteinase (CCSV-MPase) activities, isolated from *Cerastes cerastes* venom. *SFET Editions, Toxins and Ion transfers, E-Book RT19*. 2011:161–164.
- Freitas MA, Geno PW, Summer LW, Cooke ME, Hudenberg SA, Ownby CL, et al: Citrate is a major component of snake venoms. *Toxicon* 1992, **30**(4):461–464.
- Bieber AL: Metal and non-protein constituents in snake venoms. In *Snake venoms*. Edited by Lee CY. Berlin: Springer; 1979:295–304.
- Mion G, Olive F, Hernandez E, Martin YN, Viellefosse S, Goyffon M: Action des venins sur la coagulation sanguine: diagnostic des syndromes hémorragiques. *Bull Soc Path Exot* 2002, **95**(3):132–138.
- Kini RM: *Venom phospholipase A₂ enzymes: structure, function and mechanism*. England: Chichester; 1997:1–511.
- Chérifi F, Rousselle JC, Namane A, Laraba-Djebari F: CCSV-MPase, a novel procoagulant metalloproteinase from *Cerastes cerastes* venom: purification, biochemical characterization and protein identification. *Protein J* 2010, **29**:466–474.
- Kamiguti AS: Platelets as target of snake venom metalloproteinases. *Toxicon* 2005, **45**(8):1041–1049.
- Leonardi A, Fox JW, Trampus-Bakija A, Krizaj I: Ammodytase, a metalloproteinase from *Vipera ammodytes ammodytes* venom, possesses strong fibrinolytic activity. *Toxicon* 2007, **49**(6):833–842.
- Swenson S, Markland FS Jr: Snake venom fibrin(ogen)olytic enzymes. *Toxicon* 2005, **45**(8):1021–1039.
- Hutton RA, Warrel DA: Action of snake venom components on the haemostatic system. *Blood Rev* 1993, **7**(3):840–855.
- Chippaux JP, Lang J, Amadi-Eddine S, Fagot P, Le Mener V: Short report: treatment of snake envenomations by a new polyvalent antivenom composed for highly purified F(ab')₂. Results of a clinical trial in northern Cameroon. *Am J Trop Med Hyg* 1999, **61**(6):1017–1018.
- Hawgood B: Doctor Albert Calmette 1863–1933: founder of antivenom serotherapy and of antituberculous BCG vaccination. *Toxicon* 1999, **37**(9):1241–1258.
- Krifi MN, El AM, Dellagi K: The improvement and standardization of antivenom production in developing countries: comparing antivenom quality, therapeutic efficiency, and cost. *J Venom Anim Toxins* 1999, **5**(2):128–141.
- Ferreira Junior RS, Nascimento N, Couto R, Alves JB, Meira DA, Barraviera B: Laboratory evaluation of young ovines inoculated with natural or 60Co-irradiated *Crotalus durissus terrificus* venom during hyperimmunization process. *J Venom Anim Toxins incl Trop Dis* 2006, **12**(4):620–631.
- Oussedik-Oumehdi H, Laraba-Djebari F: Irradiated *Cerastes cerastes* venom as a novel tool for immunotherapy. *Immunopharmacol Immunotoxicol* 2008, **30**(1):37–52.
- Chippaux JP, Goyffon M: Venoms, antivenoms and immunotherapy. *Toxicon* 1998, **36**(6):823–846.
- Morais VM, Massaldi H: Snake antivenoms: adverse reactions and production technology. *J Venom Anim Toxins* 2009, **15**(1):2–18.
- Theakston RD, Warrell DA, Griffiths E: Report of a WHO workshop on the standardization and control of antivenoms. *Toxicon* 2003, **41**(5):541–557.
- Wilde H, Thipkong P, Sitprija V, Chaiyabutr N: Heterologous antisera and antivenins are essential biologicals: perspectives on a worldwide crisis. *Ann Intern Med* 1996, **125**(3):233–236.
- Laraba-Djebari F, Martin-Eauclaire MF: Purification and characterization of a phospholipase A₂ from *Cerastes cerastes* (horn viper) snake venom. *Toxicon* 1990, **28**(6):637–646.
- Laraba-Djebari F, Martin-Eauclaire MF, Marchot PA: A fibrinogen-clotting serine proteinase from *Cerastes cerastes* (horned viper) with arginine -

- esterase an amidase activities, purification, characterization and kinetic parameter determination. *Toxicon* 1992, **30**(1):1399–1410.
33. Chérifi F, Laraba-Djebari F: *Purification et caractérisation d'une fraction anti-coagulante et protéolytique du venin de Cerastes cerastes. Rencontres en Toxinologie, «Toxines émergentes: nouveaux risques» Lavoisier, Editions TEC et DOC.* 2007:234–235.
34. Chérifi F, Laraba-Djebari F: *Mise en évidence et Caractérisation d'une Fraction Coagulante et Agrégante du venin de Cerastes cerastes. Toxines et fonctions cholinergiques neuronales et non neuronales. Editions de la SFET.* 2008:153–154.
35. Boukhalfa-Abib H, Meksem A, Laraba-Djebari F: *Purification and biochemical characterization of a novel hemorrhagic metalloproteinase from horned viper (Cerastes cerastes) venom. Comp Biochem Physiol C Toxicol Pharmacol* 2009, **150**(2):285–290.
36. El-Asmar MF, Shaban E, Hagag M, Swelam N, Tu A: *Coagulant component in Cerastes cerastes (Egyptian sand viper) venom. Toxicon* 1986, **24**(11–12):1037–1044.
37. Bazaa A, Marrakchi N, El Ayeb M, Sanz L, Calvete JJ: *Snake venomomics: comparative analysis of the venom proteomes of the Tunisian snakes Cerastes cerastes, Cerastes vipera and Macrovipera lebetina. Proteomics* 2005, **5**(16):4223–4235.
38. Retzios AD, Markland FS: *Fibrinolytic enzymes from the venoms of Agkistrodon contortrix contortrix and Crotalus basiliscus basiliscus: cleavage site specificity towards the alpha-chain of fibrin. Thromb Res* 1994, **74**(4):355–367.
39. Hahn BS, Chang IM, Kim YS: *Purification and characterization of piscivorase I and II, the fibrinolytic enzymes from eastern cottonmouth moccasin venom (Agkistrodon piscivorus piscivorus). Toxicon* 1995, **33**(7):929–941.
40. Siigur E, Samel M, Tõnismagi K, Subbi J, Siigur J, Tu AT: *Biochemical characterization of lebetase, a direct-acting fibrinolytic enzyme from Vipera lebetina snake venom. Thromb Res* 1998, **90**(1):39–49.
41. Rodrigues VM, Soares AM, Guerra-Sá R, Rodrigues V, Fontes MR, Giglio JR: *Structural and functional characterization of neuwiedase, a non-hemorrhagic fibrin(ogen)olytic metalloprotease from Bothrops neuwiedi snake venom. Arch Biochem Biophys* 2000, **381**(2):213–224.
42. Bello CA, Hermogenes AL, Magalhaes A, Veiga SS, Gremski LH, Richardson M, et al: *Isolation and biochemical characterization of a fibrinolytic proteinase from Bothrops leucurus (white-tailed Jararaca) snake venom. Biochimie* 2006, **88**(2):189–200.
43. de Oliveira DG, Murakami MT, Cintra ACO, Franco JJ, Sampaio SV, Arni RK: *Functional and structural analysis of two fibrinogen-activating enzymes isolated from the venoms of Crotalus durissus terrificus and Crotalus durissus collilineatus. Acta Biochim Biophys Sin (Shanghai).* 2009, **41**(1):21–29.
44. Costa FL, Rodrigues RS, Izidoro LF, Menaldo DL, Hamaguchi A, Homsí-Brandeburgo MI, et al: *Biochemical and functional properties of a thrombin-like enzyme isolated from Bothrops pauloensis snake venom. Toxicon* 2009, **54**(6):725–735.
45. Zouari-Kessentini R, Luis-José L, Karray A, Kallech-Ziri O, Srairi-Abid N, Bazaa A, et al: *Two purified and characterized phospholipases A2 from Cerastes cerastes venom, that inhibits cancerous cell adhesion and migration. Toxicon* 2009, **53**(4):444–453.
46. Faure G, Gowda VT, Maroun RC: *Characterization of a human coagulation factor Xa-binding site on Viperidae snake venom phospholipases A₂ by affinity binding studies and molecular bioinformatics. BMC Struct Biol* 2007, **7**:82.
47. Franceschi A, Rucavado A, Mora N, Gutiérrez JM: *Purification and characterization of BaH4, a haemorrhagic metalloproteinase from the venom of snake Bothrops asper. Toxicon* 2000, **38**(1):63–77.
48. Marrakchi N, Barbouche R, Guermazi S, Bon C, el Ayeb M: *Procoagulant and platelet-aggregating properties of Cerastocytin from Cerastes cerastes venom. Toxicon* 1997, **35**(2):261–272.

doi:10.1186/1678-9199-19-11

Cite this article as: Chérifi and Laraba-Djebari: Isolated biomolecules of pharmacological interest in hemostasis from *Cerastes cerastes* venom. *Journal of Venomous Animals and Toxins including Tropical Diseases* 2013 **19**:11.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

